MINISTERIO DE CIENCIA E INNOVACIÓN

Cofinanciado por la Unión Europea

AGENCIA ESTATAL DE INVESTIGACIÓN

CURRICULUM VITAE ABREVIADO (CVA)

Part A. PERSONAL INFORMATION

First name	Víctor		
Family name	Jiménez Cid		
Gender (*)	Male	Birth date	
ID number			
e-mail	vicjcid@ucm.es	URL: https://www.ucm.es/signalyeast/	
Open Researcher and Contributor ID (ORCID) (*)		0000-0002-3933-2016	

(*) Mandatory

A.1. Current position

Position	Professor (Catedrático de Universidad)		
Initial date	13/12/2018		
Institution	Universidad Complutense de Madrid (UCM)		
Department/Center	Dpto. de Microbiología y Parasitología, Facultad de Farmacia		
Country	Spain	Tel.	91 3941888
Key words	Yeast, *Saccharomyces cerevisiae*, cell signaling, microbial models		

A.2. Previous positions (research activity interruptions, indicate total months)

Period	Position/Institution/Country/Interruption cause
2003-2018	Associate Professor (Profesor Titular, UCM, Spain)
2000-2003	Assistant Professor (Profesor Asociado, UCM, Spain)
1999-2000	Fulbright Postdoctoral Fellow (UC Berkeley, California, USA)
1996-2000	Assistant Professor (Profesor Ayudante Doctor, UCM, Spain)
1995-1996	Assistant Professor (Profesor Ayudante, UCM, Spain)
1992-1995	Graduate Researcher (F.P.U. PhD Student, UCM, Spain)

A.3. Education

PhD, Licensed, Graduate	University/Country	Year
PhD in Pharmacy	Universidad Complutense de Madrid (UCM)	1996
Degree in Pharmacy	Universidad Complutense de Madrid (UCM)	1991

Part B. CV SUMMARY *(max. 5000 characters, including spaces)*

Graduated in Pharmacy on 13/09/1991, average Bachelor's Grade 9.43/10; Graduation Award "**Premio Extraordinario Complutense de Licenciatura y Diplomatura 1990-91, area Ciencias Biomédicas"**. A FPU Fellowship supported my **PhD in Pharmacy**, (05/07/1996) with *cum laude* qualification. After a Postdoc at the University of California at Berkeley in the prestigious Dr. Thorner´s lab, supported by Fulbright and Del Amo Fellowships, got on a tenure track on **Microbiology** as Assistant Professor (05/02/2003). In July 2015 I was accredited by the National Evaluation Committee (ANECA) for Full Professor (**Catedrático**), position that I enjoy from Dec. 2018. I have four research 6-year positive evaluations ("sexenios"), currently applying for a 5th one, and five 5-year teaching periods ("quinquenios"). In 2022, I was nominated as Corresponding Academic of the *Real Academia Nacional de Farmacia*, and Academic (medal 26) of the *Real Academia de Doctores de España*. I have taught Microbiology-related credits in several Degrees and Masters obtaining "**Positive**" or "**Very Positive**" assessments by the "Docentia" program, based on student surveys, in 19 occasions. I directed courses on SARS-CoV-2 in the UCM Summer School at San Lorenzo de El Escorial (2000, 2021).

As a researcher, I have participated in over **20 Research Projects**, being principal investigator of the ongoing PID2019-105342GB-I00 Grant, which I co-direct with Prof. Maria Molina. The indicators of my scientific production can be consulted in SCOPUS (ID 6603604065; h-index 25, 2103 citations) or Google Scholar (h-index 29, 3117 citations, i10 index 46). I have published **59 articles** in JCR indexed journals, 63% of them in relevant Q1

journals in the areas of Microbiology, Molecular Biology or Genetics, 58 % as first, last or corresponding author. My research is focused on the study of cell signaling in the eukaryotic model yeast *Saccharomyces cerevisiae* and its exploitation to study heterologous genes related to human disease, namely microbial virulence factors, cancer, innate immunity and inflammation, a field that our group is pioneering. Highlights in my early scientific career were my seminal contributions to morphogenetic regulation in *S. cerevisiae* (Cid *et al*., 1998, *Microbiology* doi: 10.1099/00221287-144-12-3463; Jimenez *et al*. 1998 *J Cell Biol* doi: 10.1083/jcb.143.6.1617), the elucidation of molecular mechanisms of the morphogenetic cell cycle checkpoint (Cid *et al*., 2001 *Mol Biol Cell*. doi: 10.1091/mbc.12.6.1645), the discovery of molecular mechanisms host cell signaling subversion by *Salmonella* using the yeast model (Rodríguez-Escudero *et al*. 2011. *Mol Microbiol*. doi: 10.1111/j.1365-2958.2011.07639.x), and the development of a yeast-based platform to study the oncogenic PI3K/PTEN/Akt pathway in collaboration with Dr. R. Pulido (Rodríguez-Escudero *et al*., 2005. *Biochem J*. doi: 10.1042/BJ20050574, Cid *et al*., 2008. *Oncogene*. doi: 10.1038/onc.2008.240). These were the basis of my subsequent trajectory in this field. I currently enjoy several **international collaborations**, established after stays as Visiting Scholar at the J. Kagan lab (Boston Children's Hospital, Harvard Medical School, June-Sept. 2016), and at the J. Thorner lab (UC Berkeley, June-Aug. 2018). My current interest is the development of humanized yeast systems for molecular studies on inflammatory disease and the pathogenicity of infectious diseases. I have directed **6 PhD Thesis**, and 2 more will be defended shortly.

Compromised with Communication of Science and social outreach, I was one of the founding members of the Communication division of the **Spanish Society for Microbiology** (SEM), a scientific Society in which I am involved from 2012 as member of the Steering Committee, first as Editor of the members' magazine, SEM@foro (2012-2018) and later as Treasurer (2018-now). From SEM, I coordinate the **MicroMundo** project, a **Citizen Science and Service-Learning** project focused on **antibiotic resistance awareness**, partner of the Small World Initiative/Tiny Earth projects in the US, operating in 30 Universities in Spain and Portugal, and awarded **national and international prizes**: FundaMed (Spain, 2018), Antibiotic Guardian (UK, 2019) and PRAN (Spain, 2021).
During the SARS-CoV-2 pandemics, I have actively participated in **science communication** by collaborating with mass media (public and private live TV broadcasts, radio, digital newspapers, blogs –the Conversation, podcasts –Twitch, etc.).
I was a member of the FEMS (Federation of European Microbiology Societies) Grants Committee from 2017 to 2022, and have been a referee in panels of diverse National and International calls for research grants. I have been Editor or member of the Editorial Board of several journals: Microbiology (2009-2019), Microbiology Open (2011-2019), Genes (2018-now) and Scientific Reports (2019-now).

Part C. RELEVANT MERITS *(sorted by typology)*

C.1. Publications. *(CA) stands for Corresponding Author.*
I list here the **10 articles from the last 5 years** that are most relevant for the project, i. e., focused on humanized yeast, cell signaling and molecular mechanisms of human disease:

1. Valenti M, Molina M, **Cid VJ (CA)**. 2023. Human Gasdermin D and MLKL disrupt mitochondria, endocytic traffic and TORC1 signaling in budding yeast. *Open Biol.* 13: 220366. doi.org/10.1098/rsob.220366. IF:7.000 (Q1)
2. Sá-Pessoa J, López-Montesino S, (…), **Cid VJ (11/12)**, Bengoechea JA. 2023. A trans-kingdom T6SS effector targets the mitochondria activating the innate receptor NLRX1 to promote infection. *Nature Commun.* doi:10.1038/s41467-023-36629-3. IF 17.694 (Q1).
3. Torices L, (…), **Cid VJ (12/13)**, Pulido R. 2022. Functional analysis of PTEN variants of unknown significance from PHTS patients unveils complex patterns of PTEN biological activity in disease. *Eur J Hum Genet.* doi: 10.1038/s41431-022-01265-w. IF 5.351 (Q1).
4. Coronas-Serna JM, del Val E, Kagan JC, Molina M, **Cid VJ (CA).** 2021. Heterologous Expression and Assembly of Human TLR Signaling Components in *Saccharomyces cerevisiae*. *Biomolecules*. 11:1737. doi: 10.3390/biom11111737. IF: 6.064 (Q2).
5. Valenti M, Molina M, **Cid VJ (CA)**. Heterologous Expression and Auto-Activation of Human Pro-Inflammatory Caspase-1 in *Saccharomyces cerevisiae* and Comparison to Caspase-8. Front Immunol. 2021 12:668602. doi: 10.3389/fimmu.2021.668602. IF 8.787(Q1).

6. Luna S, (…), **Cid VJ (13/14)**, Pulido R. 2021. A global analysis of the reconstitution of PTEN function by translational readthrough of PTEN pathogenic premature termination codons. *Hum Mutat*. 42:551-566. doi: 10.1002/humu.24186. IF.4.7 (Q2).
7. Coronas-Serna JM, (…), **Cid VJ (10/11; CA)**, Salcedo SP. 2020. The TIR-domain containing effectors BtpA and BtpB from *Brucella abortus* impact NAD metabolism. *PLoS Pathog*. 16(4):e1007979. doi: 10.1371/journal.ppat.1007979. IF 6.823 (Q1).
8. Storey D, (…), **Cid VJ (10/12)**, Salminen TA, Bengoechea JA. 2020. *Klebsiella pneumoniae* type VI secretion system-mediated microbial competition is PhoPQ controlled and reactive oxygen species dependent. *PLoS Pathog*. 16(3):e1007969. doi: 10.1371/journal.ppat.1007969. IF 6.823 (Q1).
9. Coronas-Serna JM, Fernández-Acero T, Molina M, **Cid VJ (CA).** 2018. A humanized yeast-based toolkit for monitoring phosphatidylinositol 3-kinase activity at both single cell and population levels. *Microb Cell*. 5:545-554. doi: 10.15698/mic2018.12.660. IF (2021, 1st year in JCR) 5.316 (Q2).
10. Rodríguez-Escudero I, Fernández-Acero T, **Cid VJ (CA),** Molina M. 2018 Heterologous mammalian Akt disrupts plasma membrane homeostasis by taking over TORC2 signaling in *S. cerevisiae*. *Sci Rep*. 8:7732. doi:10.1038/s41598-018-25717-w. IF: 4.011 (Q1).

C.2. Congresses, indicating the modality of their participation (invited conference, oral presentation, poster)

I have presented over **100 communications to National and International Congresses**. Some recent communications related to the topic of the Project are listed here.

1. El domino N-terminal de la gasdermina D provoca alteraciones en el ciclo celular, el tráfico endosomal y las mitocondrias en *Saccharomyces cerevisiae.* Valenti M, Molina M, **Cid VJ.** XIII Reunión Nacional Microbiología Molecular. Granada, 7-9 sept. 2022. Oral Communication.
2. Heterologous expression and assembly in *Saccharomyces cerevisiae* of Death Domain-containing components of human myddosome. del Val E, Coronas-Serna JM, **Cid VJ,** Molina M. EMBL Symposium "Innate Immunity in host-pathogen interactions", Heidelberg, Germany, 17-20 July 2022. Poster.
3. Heterologous Expression and Assembly in Saccharomyces cerevisiae of Death Domain-containing Components from Human TLR-signaling. del Val E, Coronas-Serna JM, Molina M, **Cid VJ**. World Microbe Forum 2021. 20-24 June 2021. Online. Poster.
4. Heterologous expression of human pro-inflammatory Caspase-1 in Saccharomyces cerevisiae and comparison to pro-apoptotic Caspase-8. Valenti M, Molina M, **Cid VJ**. World Microbe Forum 2021. 20-24 June 2021. Online. Poster.
5. Humanized Yeast Models in Biomedicine: Assembling Signaling Modules in Cancer and Innate Immunity. del Val E, Valenti M, Coronas-Serna JM, Molina M, **Cid VJ**. Yeast in the intrersection between Systems Biology and Biomedicine. Fundación Ramón Areces Symposium. Madrid, 23-24 Jan. 2020. Invited Conference.
6. Humanized yeast models: *Saccharomyces cerevisiae* as a tool on research on the oncogenic PI3K pathway Coronas-Serna JM, del Val E, Valenti M, Fernandez-Acero T, Rodriguez-Escudero I, Molina M, **Cid VJ.** World Yeast Congress. Vancouver, Canada July 26-27 2019. Oral Communication.
7. *Brucella abortus* TIR domain-containing effectors are toxic in *Saccharomyces cerevisiae* due to depletion of NAD^+. Coronas-Serna JM, Louche A, Rodríguez-Escudero M, Rodríguez-Escudero I. Molina M, Salcedo S, **Cid VJ.** ASM Microbe, San Francisco, USA, 20 - 24 Jun 2019. Oral Communication.
8. Levaduras humanizadas: diseñando modelos para estudios moleculares en patologías hereditarias. **Cid VJ**., Coronas-Serna JM, del Val E, Fernández-Acero T, Rodríguez-Escudero I, Molina M. XIV Congreso Nacional de Micología. Tarragona, 17-19/09/2018. Invited conference.
9. In search of novel toolkits to study TLR signaling: modular assembly of myddosome components in an heterologous yeast model. **Cid VJ**, Coronas-Serna JM, del Val E, Genna G, Fernández-Acero T, Kagan J, Molina M. EMBO-EMBL Symposium: Innate Immunity in Host-Pathogen Interactions. Heidelberg, Germany, 9-11/06/2018. Poster.
10. *Saccharomyces cerevisiae* as a Model to Study TIR-Domain Containing Proteins in Host-Pathogen Interactions. Coronas-Serna JM, Fuentes V, Rodríguez-Escudero I; Molina M.

Kagan J, Salcedo SP, **Cid VJ.** 28th International Conference on Yeast Genetics and Molecular Biology (ICYGMB) Prague, Czech Republic, 27/08/2017: Poster.

C.3. Research projects, indicating your personal contribution. In the case of young researchers, indicate lines of research for which they have been responsible.
I list here the research projects I have been involved along in the **latest 10 years**, which have supported the consolidated line of research that relates to this project. Besides these, I have been **Principal Investigator** of the FECYT project **FCT-17-12215** ("SWI@Spain: Implantación y consolidación en España de Small World Initiative, un proyecto de Ciencia Ciudadana para el uso racional y descubrimiento de nuevos antibióticos").

PID2019-105342GB-I00. "Ensamblaje y estudio de complejos de señalización endógenos y heterólogos en el modelo *Saccharomyces cerevisiae*: aplicación al análisis funcional y descubrimiento de fármacos." **Principal Investigator.**
Funding Agency: Ministerio de Ciencia e Innovación. Funding: 193,600.00€.
Principal investigators: María Molina y **Víctor J. Cid**. Dates: 01-06-2020 - 31-05-2023

S2017/BMD-3691. InGEMICS-CM: Ingeniería Microbiana, Salud y Calidad de Vida.
Funding Agency: Comunidad Autónoma de Madrid. AYUDAS A GRUPOS PARA EL DESARROLLO DE PROGRAMAS DE ACTIVIDADES DE I+D EN BIOMEDICINA. **Researcher.** Dates: 1-1-2018 al 31-12-2022. Total funding: 1,037,199 €. Subproject SIGNALYEAST IP: María Molina, Fund.: 161,512.73 €.

BIO2016-75030-P. "Reprogramación celular por fosforilación dependiente de la MAPK SLT2 e integración de un módulo de señalización por receptores de tipo Toll en *S. cerevisiae*". **Researcher.** Dates 30/12/2016 - 31/12/2020
Funding Agency: Ministerio de Economía y Competitividad. Funding: 190,000.00 €
Principal Investigators: María Molina y Humberto Martin Brieva.

PIM2010EPA-00664. ERA-NET PathoGenoMics. "Caracterizacion de rutas del hospedador alteradas por efectores de *Brucella, Chlamydia* y *Coxiella*".
Funding Agency: Programa Nacional de Internacionalización de la I+D. SUBPROGRAMA: PROYECTOS INTERNACIONALES. MODALIDAD A - Proyectos Multilaterales. **Researcher.** Dates 01/03/2011 al 30/06/2014 Funding: 100,000 €
Coordinator: Jaime Mota; Principal Investigator UCM: Isabel Rodríguez Escudero

BIO2013-44112-P "Identificación de nuevos componentes y reconfiguración de circuitos de señalización en *Saccharomyces cerevisiae*¨. **Researcher.**
Funding agency: Ministerio de Economía y Competitividad. Dates l 01/01/2014- 31/12/2017
Principal Investigator: María Molina. Funding 180,000.00 €

PROMPT S2010/BMD-2414. "Programación de circuitos microbianos en medicina protectiva y terapeútica". **Researcher.** Dates:1/1/2012 - 30/4/2016
Funding Agency: Comunidad de Madrid. Funding 670,951.99 €
Subproject Principal Investigator: María Molina Funding 141,740,00 €

BIO2010-22369. "*S. cerevisiae* como modelo para el estudio de la señalización eucariótica y su aplicación al diseño de bioensayos de cribado farmacológico". **Researcher.**
Funding Agency: Ministerio de Ciencia e Innovación. Funding: 200.000,00 €
Principal Investigator: María Molina. Dates: 01/01/2011 al 30/06/2014

C.4. Contracts, technological or transfer merits, Include patents and other industrial or intellectual property activities (contracts, licenses, agreements, etc.) in which you have collaborated. Indicate: a) the order of signature of authors; b) reference; c) title; d) priority countries; e) date; f) Entity and companies that exploit the patent or similar information, if any

WO 2016/128401. Compounds to treat cancer.
De Pedro N, González-Menéndez V, Crespo G, Cautain B, Fernández-Acero T, **Cid VJ,** Molina M., Vicente F, Reyes JF, Martín-Serrano JM, Perez-Victoria I, Genilloud O (collaboration with Fundación MEDINA, Granada). Date: 07/04/2015. International Patent. Not exploited.